

Comparison of Histological Changes in the Ligamentum Flavum from Spinal Canal Stenosis Patients with and without Diabetes Mellitus

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Abstract

Objective: Lumbar spinal canal stenosis (LSCS) has prominent position in spinal disorders of the elderly. Ligamentum flavum hypertrophy plays a dominant role in narrowing of the LSCS. There are some clues that Ligamentum flavum hypertrophy is associated with diabetes mellitus. Our objective was comparison of histological properties of ligamentum flavum in Lumbar spinal canal stenosis patients with and without diabetes mellitus.

Materials and Methods: In this case control study, during 9 months, twenty-nine patients, who were candidate for decompressive laminectomy because of ligamentum flavum hypertrophy, were studied. They were labeled as diabetic if had history of diabetes mellitus or fasting blood sugar ≥ 126 mg/dl for two times or blood sugar >200 mg/dl anytime with symptom of hyperglycemia. Ligamentum flavum was removed through surgery. Samples were evaluated by pathologist using Hematoxylin & Eosin, Masson's trichrome and Verhoeff van Gieson staining. Evaluation includes grading of fibrosis, loss of elastin fibers, calcification and number of cellularity of samples. Data analyzed with Fisher's test and Mann-Whitney test.

Results: In this study 41% of patients (12 persons) were suffering from diabetes mellitus. There was no significant difference between diabetic and nondiabetic patients in histological properties. ($P>0.05$); but the weight of diabetic patients was higher than nondiabetics (P -value =0.038)

Conclusion: These results show that diabetes mellitus has no effect on histological change in ligamentum flavum of spinal canal stenosis and mechanical stress (due to overweight) has a more important role in pathogenesis of spinal canal stenosis.

Keywords: Diabetes mellitus, Spinal canal stenosis, Ligamentum flavum, Histological change, Overweight, Tumor growth factor- β

Introduction

Lumbar spinal canal stenosis (LSCS) is one of the most common spinal disorders in the elderly. The causes of LSCS include ligamentum flavum (LF) hypertrophy, hypertrophy of the facet joints, bulging of the intervertebral discs, and vertebral endplate osteophytosis. LF hypertrophy plays a dominant role in the

narrowing of the lumbar spinal canal (1). Fibrosis is considered as the main cause of LF hypertrophy. Transforming growth factor (TGF- β) released by endothelial cells may stimulate the fibrosis, especially during the early phase of hypertrophy (2). The structure of LF is unique, because of a predominance of elastic fibers, and its intrinsic innervations at each level of the spine. The innervations decrease with increasing degeneration (3). Histological changes in the hypertrophied LF from LSCS patients include fibrosis, degradation of elastic fibers with an increase in collagen fibers, granulation tissue proliferation, chondroid metaplasia, and calcification (2,4,5) increasing of TGF- β (2,6,7,8). Although the serum concentration of TGF- β was normal (9). In the other hand TGF- β has an important role in complications of diabetes mellitus. TGF- β is increased in diabetic nephropathy and stimulates basement membrane production of collagen and fibronectin by mesangial cells (10). In some articles, mechanical stress introduced as the increasing of TGF- β cause (2,11,6). In the other study, it was seen that expression of MMP13, which is involved in inflammation and fibrosis, in the LF from lumbar canal stenosis patients with diabetes mellitus was higher than non-diabetic patient (12). It has mentioned that diabetes mellitus is a risk factor for the development of lumbar spinal stenosis (13).

Our purpose was to compare the histological changes (fibrosis, loss of elastin, calcification and cellularity) in LF samples from the patients with spinal canal stenosis between the diabetic and non-diabetic groups.

Materials and Methods

In this study, from April 2013 up to 9 months we studied all the patients that get under decompressive laminectomy because of hypertrophy of LF in Shahid Rahnemoun Hospital. The inclusion criteria was, LF hypertrophy that was diagnosed by neurosurgeon in the lumbar MRI. Patients, who had spinal canal stenosis because of

tuberculosis, ossification of posterior longitudinal ligament (OPLL), vertebral fracture and spondylolisthesis were excluded. Twenty-nine patients were studied. They were diabetic if had history of diabetes mellitus or fasting blood sugar ≥ 126 mg/dl for two times or random blood sugar > 200 mg/dl anytime with symptom of hyperglycemia. According to this definition 12 people have diabetes mellitus. First time, FBS was checked in the hospital and for second time after discharging. LF was removed during surgery. All surgeries were done in Shahid Rahnemoun hospital in Yazd, Iran. In the operation, LF was removed and immediately was placed in formalin 4% solution. Then paraffin blocks was prepared from samples and got sectioned for hematoxylin & eosin, Masson's trichrome and Verhoeff van Gieson staining.

In next step, prepared sections get evaluated about grading of fibrosis, elastin fiber decrease, calcification and number of cellularity. According to these point 5 fields was selected randomly from central (not dorsal or dural) with the $\times 40$ magnification and this four factors determined by a pathologist. Gradings were selected according to table 1 and figure 1. In order to determine calcification, the minimum amount of calcification was assumed as + (figure 1-A) and the maximum amount of calcification was assumed as +++ (figure 1-B) and the amount of calcification which was among them as ++ (figure 1-C). Data input in SPSS17 and analyzed with Fisher's exact test and Mann-Whitney test.

Results

Totally 29 patients were studied. Demographic characteristics of patients is presented in table 2.

The cellularity average in five fields and the amount of calcification in LF samples were determined; but there were no significant differences between diabetic and non-diabetics. (Figures 2-3)

Branching and variability of sizes in elastin fibers which had been placed in margin of

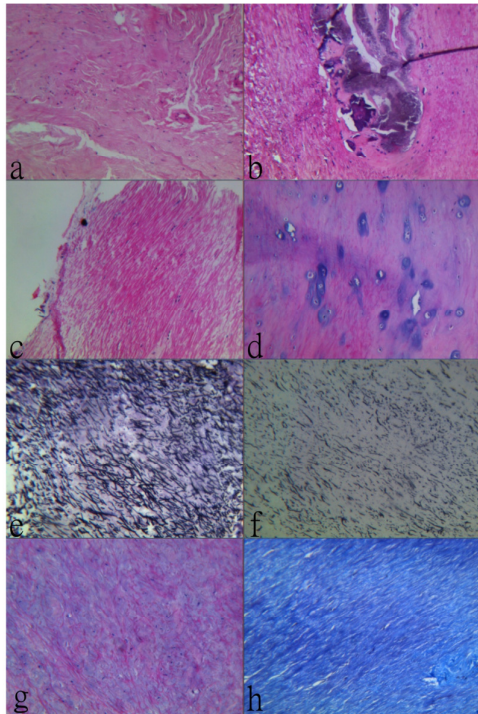


Figure 1. section without calcification stained by H&E (a) section with +++ calcification stained by H&E (b) section with + calcification stained by H&E (c) section demonstrate fibrocartilage tissue (d) section demonstrate grade0 in loss of elastin fibers stained by Verhoeff van Gieson (e) section demonstrate grade1 in loss of elastin fibers stained by Verhoeff van Gieson (f) section demonstrate grade0 in fibrosis stained by Masson's trichrome (g) section demonstrate grade1 in fibrosis stained by Masson's trichrome(h)

fibrotic region, was increased. (Figure 4).there wasn't significant difference in fibrosis grading between diabetics and non-diabetics. (Figure 5)

Conclusion

Lumbar spinal canal stenosis (LSCS) is one of the most common spinal disorders in the elderly (12). Since 1913 that Elsberg first has reported the case showing sciatica caused by the LF hypertrophy, there are several attempts to clarify the pathomechanism of LF hypertrophy but it is still unclear (2). It was reported that in lumbar canal stenosis unique histological changes are seen including, increase of collagen fibers (fibrosis) (2,4,12,14,15,16), loss of elastic fibers (2,12,15,16), calcification (4,16) and increase of cellularity (11). Decreased elastin fibers and increased collagen fibers have several reasons: aging (2), inflammation (14) and mechanical stress (6,11,17). Fibrosis is the main cause of LF hypertrophy, and TGF- β released by endothelial cells, may stimulate the fibrosis, especially during the early phase of hypertrophy (2). Nakatani et al (6) confirmed the effects of TGF- β application on collagen synthesis of cultured LF fibroblasts. It was reported that TGF- β mediated by mechanical stress induces collagen synthesis on a variety of cells in vitro (6,18,19,20). There are documents that can interpret high expression of TGF- β in LF: 1-Mechanical stress 2-diabetes mellitus.

In some studies, mechanical stress introduced as the cause of TGF- β increasing (2,6,11); so the dorsal side of LF was subjected to more stress when compared to the dural side so dorsal side of LF has more fibrotic region than dural side (2).

TGF- β has an important role in complications

Table 1. Grading of fibrosis, elastin fiber decrease, calcification

Variable	Fibrosis	Loss of elastin fibers	Calcification
Grade 0	Fibrosis at $\leq 50\%$ of all area	Elastin at $\leq 50\%$ of all area	0 and +
Grade 1	Fibrosis at $> 50\%$ of all area	Elastin at $> 50\%$ of all area	++ and +++

Table 2. Demographic data of patients

Variable	Controls	Diabetic patients
Number	17 (59%)	12 (41%)
Age	54.8 \pm 12.3	61.2 \pm 9.5
Sex		
Male	6 (35%)	8 (65%)
Female	11 (65%)	4 (35%)
Metabolic syndrome	8 (47%)	10 (83%)
BMI	25.7 \pm 4.8	26.2 \pm 8.0
Weight	67.0 \pm 10.0	72.0 \pm 18.1

of diabetes mellitus. TGF- β is increased in diabetic nephropathy and stimulates basement membrane production of collagen and fibronectin by mesangial cells (10). High blood glucose could promote collagen synthesis in the posterior longitudinal ligament mainly via endogenous TGF- β 1, resulting in hypertrophy of ligament (21). This fact supports this hypothesis that TGF- β in LF can be attributed to diabetes mellitus.

Diabetes mellitus introduced as a risk factor of several soft tissue disorders such as ossification of posterior longitudinal ligament (OPLL) (21), Achilles tendon thickness (22,23), plantar fasciitis (22), plantar

aponeurosis thickness (24), flexor hallucis longus tendon thickness (24), supraspinatus tendon thickness (25), biceps tendon thickness (25), adhesive capsulitis (26) and diffused idiopathic skeletal hyperostosis (DISH) (27). But there are a few numbers of studies about the effect of DM on LF. Fibrosis is the major change in LF of spinal canal stenosis patients (2), so if diabetes mellitus become the cause of spinal canal stenosis, different histological difference of histological change between diabetic and non-diabetic patients. In order to this point, we compare the histological change, include fibrosis, loss of elastin fibers, calcification and cellularity between the

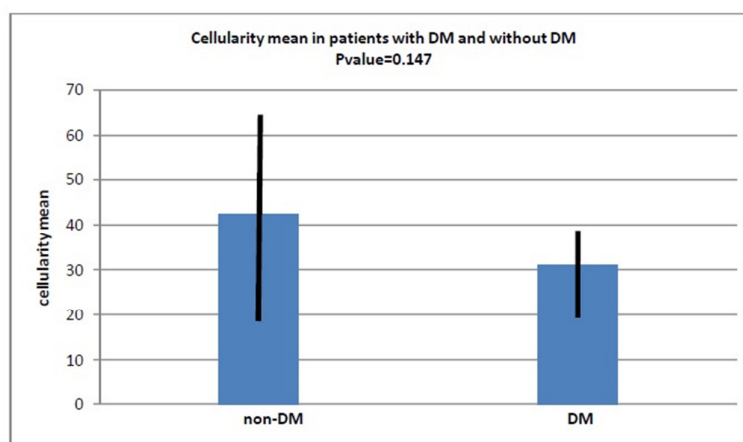


Figure 2. Cellularity mean in diabetic and non-diabetic patients. *P*-value=0.147

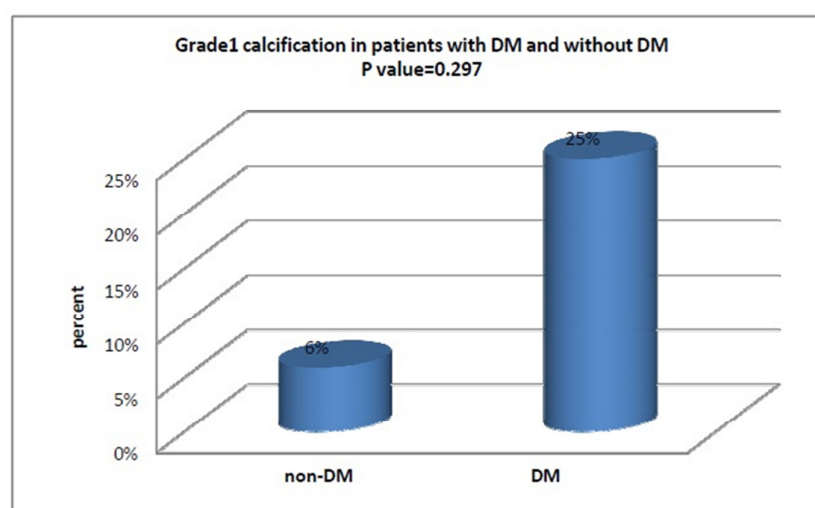


Figure 3. Grade1 calcification in diabetic and non-diabetic patients. *P*-value=0.297

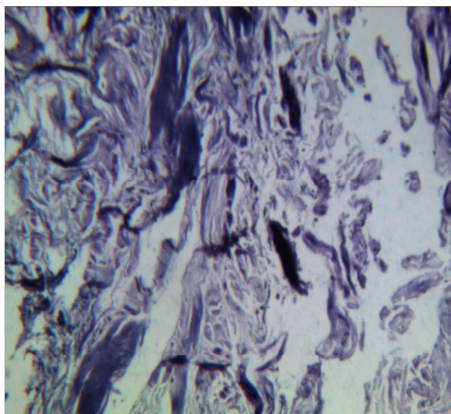


Figure 4. Disorganization, branching and variability of sizes in elastin fibers

samples of diabetic and non-diabetic patients but as it was mentioned, there was no significant difference between them. It showed that diabetes mellitus cannot be assumed as a risk factor for spinal canal stenosis.

It is supported with this fact that there was no significant difference in onset age of back pain and even diabetic patients get symptomatic in higher age (61.2 ± 9.5 vs. 54.8 ± 12.3).

On the other hand, it was known that diabetic patients are heavier than non-diabetic patient and a BMI of ≥ 25 kg/m² is a risk factor for type2 DM (10).

It is seen in our result, too. The mean of weight in diabetic is 72.0 ± 18.1 and in non-diabetic is 67.0 ± 10.0 ($P=0.024$). It was said that Patients with a BMI of ≥ 25 kg/m² had the thickest LF at the L3–L4 level (28). High weight can result in high mechanical stress and this can explain why spinal canal stenosis is more prevalent in diabetic patients as it was said by Yoram Anekstein et al (13).

This fact was seen in our data, too. As diabetes mellitus prevalence is 16.3 percent in Yazd general population ≥ 30 y/o (29); but in this study, 41% of patients were suffering from diabetes mellitus. So diabetes mellitus cannot consider as an independent risk factor for spinal canal stenosis. As it was mentioned, in 2011 a study was done by Guanyu Cui (12) that shows high expression of MMP13 in LF of diabetic patients.

In this study, doesn't say if the weight of two groups gets matched. If their weight has significant difference between diabetic and non-diabetics, it can support this hypothesis that mechanical stress is the etiology of spinal canal stenosis; Because it was demonstrated that in other tissues, mechanical stress can induce expression of MMP13 (30,31). Overly it can say that mechanical stress has an important role in LF hypertrophy.

No previous studies have addressed the difference between histological factors of LF in diabetic and non-diabetic patients. Although our study was a unique study for evaluating the effect of DM on histological change of LF, but our study had some limitation. In this study we don't consider the duration and the severity of diabetes mellitus.

If HgbA1C of patients get checked, this pitfall can be reduced. Our data can be completed if the thickness of LF between diabetic and non-diabetic get compared based on MRI. For more reliable data it has to do this study with more patients and to follow patients for long-time. Undesirably, by processing the samples there are some errors.

Thickness of sections may differ with each other, so cellularity may be unreliable and empirical evaluation of sections isn't perfect and we have to use image interpreter software. From this study, it can be concluded that diabetes mellitus hasn't direct effect on LF of LSCS patients, but because of coherency between DM and overweight, there is high prevalence of DM in LSCS patients.

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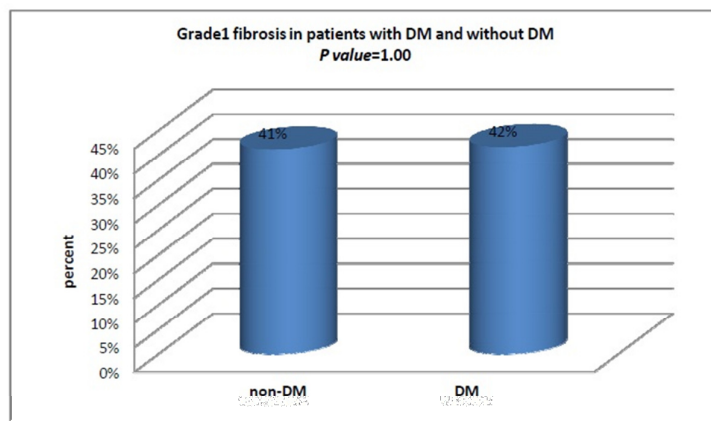


Figure 5. Grade1 fibrosis in diabetic and non-diabetic patients. *P*-value=0.90

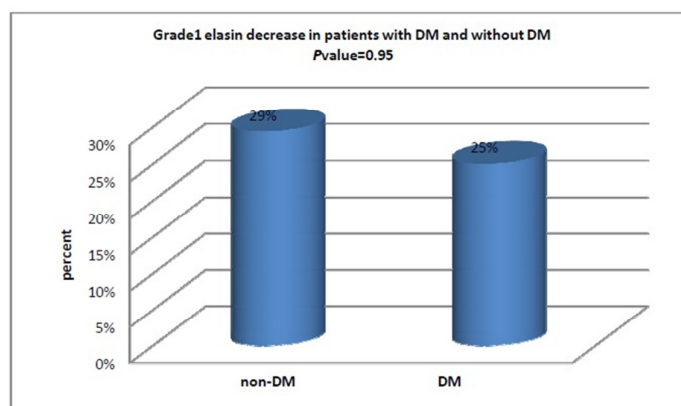


Figure 6. Grade1 elasin decrease in diabetic and non-diabetic patients. *P*-value=0.95

References

- Hansson T, Suzuki N, Hebelka H, Gaulitz A. The narrowing of the lumbar spinal canal during loaded MRI: the effects of the disc and ligamentum flavum. *Eur Spine* 2009;18:679-86.
- Sairyo K, Biyani A, Goel V, Leaman D, Booth R, Thomas J, et al. Pathomechanism of Ligamentum Flavum Hypertrophy: A Multidisciplinary Investigation Based on Clinical, Biomechanical, Histologic, and Biologic Assessments. *Spine* 2005;23(30): 2649-56.
- Viejo-Fuertes D, Liguoro D, Rivel J, Midy D, Guerin J. Morphologic and histological study of the ligamentum flavum in the thoraco-lumbar region. *Surg Radiol Anat* 1998;20:171-6.
- Schröder PK, Grob D, Rahn BA, Cordey J, Dvorak J. Histology of the ligamentum flavum in patients with degenerative lumbar spinal stenosis. *Eur Spine J* 1999;8:323-28.
- Yoshida M, Shima K, Taniguchi Y, Tamaki T, Tanaka T. Hypertrophied ligamentum flavum in lumbar spinal canal stenosis. Pathogenesis and morphologic and immunohistochemical observation. *Spine* 1992;17(11):1353-60.
- Nakatani T, Marui T, Hitora T, Doita M, Nishida K, Kurosaka M. Mechanical stretching force promotes collagen synthesis by cultured cells from human ligamentum flavum via transforming growth factor- β 1. *Journal of Orthopaedic Research* 2002;20(6):1380-86.
- Park J, Chang H, Lee J. Quantitative analysis of transforming growth factor-beta 1 in ligamentum flavum of lumbar spinal stenosis and disc herniation. *Spine* 2001;26(21): 492-5.
- Zhong Z, Chen J. Overexpression of transforming growth factor-beta1 in degenerative ligamentum flavum. *Nan Fang Yi Ke Da Xue Xue Bao* 2009;29(2):316-8.

9. Kim H, Park J, Won H, Chang H. Serum Levels of TGF-beta1, TIMP-1 and TIMP-2 in Patients with Lumbar Spinal Stenosis and Disc Herniation. *Asian Spine J* 2007;1(1):8-11.
10. Longo D, Fauci A, Kasper D, Hauser S, Jameson J, Loscalzo J. *Harrison's Principles of Internal Medicine*. Medical Publishing Division 2012;18.
11. Shafaq N, Suzuki A, Terai H, Wakitani S, Nakamura H. Cellularity and Cartilage Matrix Increased in Hypertrophied Ligamentum Flavum. *Spinal Disord Tech* 2012;25(2):107-15.
12. Cui G, Watanabe K, Miyauchi Y, Hosogane N, Tsuji T, Ishii K, et al. Matrix metalloproteinase 13 in the ligamentum flavum from lumbar spinal canal stenosis patients with and without diabetes mellitus. *J Orthop Sci* 2011;16: 785-90.
13. Anekstein Y, Smorgick Y, Lotan R, Agar G, Shalmon E, Floman Y, et al. Diabetes mellitus as a risk factor for the development of lumbar spinal stenosis. *Isr Med Assoc J* 2010;12(1):16-20.
14. Sairyo K, Biyani A, Goel V, Leaman D, Booth R, Thomas J, et al. Lumbar ligamentum flavum hypertrophy is due to accumulation of inflammation-related scar tissue. *Spine* 2007;32(11):340-7.
15. Kosaka H, Sairyo K, Biyani A, Leaman D, Yeasting R, Higashino K, et al. Pathomechanism of Loss of Elasticity and Hypertrophy of Lumbar Ligamentum Flavum in Elderly Patients With Lumbar Spinal Canal Stenosis. *Spine* 2007;32(25):2805-11.
16. Okuda T, Baba I, Fujimoto Y, Tanaka N, Sumida T, Manabe H, et al. The pathology of ligamentum flavum in degenerative lumbar disease. *Spine* 2004;29(15):1689-97.
17. Löhr M, Hampl JA, Lee JY, Ernestus RI, Deckert M, Stenzel W. Hypertrophy of the lumbar ligamentum flavum is associated with inflammation-related TGF- β expression. *Acta neurochirurgica* 2011;153(1):134-41.
18. Hirakata M, Kaname S, Chung UG, Joki N, Hori Y, Noda M, et al. Tyrosine kinase dependent expression of TGF- induced by stretch in mesangial cells. *Kidney International* 1997;51:1028-36.
19. Lee T, Chin G, Kim W, Chau D, Gittes G, Longaker M. Expression of transforming growth factor beta 1, 2, and 3 proteins in keloids. *Ann Plast Surg* 1999;43(2):179-84.
20. Yasuda T, Kondo S, Homma T, Harris R. Regulation of extracellular matrix by mechanical stress in rat glomerular mesangial cells. *J Clin Invest* 1996;98(9):1991-2000.
21. Li H, Liu D, Zhao CQ, Jiang LS, Dai LY. High glucose promotes collagen synthesis by cultured cells from rat cervical posterior longitudinal ligament via transforming growth factor-b1. *Eur Spine* 2008;17: 873-81.
22. Giacomozzi C, D'Ambrogi E, Uccioli L, Macellari V. Does the thickening of Achilles tendon and plantar fascia contribute to the alteration of diabetic foot loading? *Clin Biomech* 2005;20(5):532-9.
23. Akturk M, Ozdemir A, Maral I, Yetkin I, Arslanl M. Evaluation of Achilles Tendon Thickening in Type 2 Diabetes Mellitus. *Exp Clin Endocrinol Diabetes* 2006;115:92-96.
24. Bolton N, Smith K, Pilgram T, Mueller M, Bae K. Computed tomography to visualize and quantify the plantar aponeurosis and flexor hallucis longus tendon in the diabetic foot. *Clin Biomech* 2005;20(5):540-6.
25. Akturk M, Karaahmetoglu S, Kacar M, Muftuoglu O. Thickness of the supraspinatus and biceps tendons in diabetic patients. *Diabetes Care* 2002;25(2):408.
26. Wang K, Ho V, Hunter-Smith D, Beh P, Smith K, & Weber A. Risk factors in idiopathic adhesive capsulitis: a case control study. *J Shoulder Elbow Surg* 2013;22(7):24-9.
27. Coaccioli S, Fatati G, Di Cato L, Marioli D, Patucchi E, Pizzuti C, et al. Diffuse idiopathic skeletal hyperostosis in diabetes mellitus, impaired glucose tolerance and obesity. *Panminerva Med* 2000;42(4):247-51.
28. Altinkaya N, Yildirim T, Demir S, Alkan O, Sarica FB. Factors Associated With the Thickness of the Ligamentum Flavum. *Spine* 2011;36(16):1093-97.
29. Lotfi M, Saadati H, Afzali M. Prevalence of Diabetes in People aged ≥ 30 years: The Results of Screen-ing Program of Yazd Province, Iran, in 2012. *J Res Health Sci* 2014;14(1):87-91.
30. Yang CM, Chien CS, Yao CC, Hsiao LD, Huang YC, Wu CB. Mechanical Strain Induces Collagenase-3 (MMP-13) Expression in MC3T3-E1 Osteoblastic Cells. *The Journal of Biological Chemistry* 2004;279(21): 22158-65.
31. Leigh D, Abreu E, Derwin K. Changes in gene expression of individual matrix metalloproteinases differ in response to mechanical unloading of tendon fascicles in explant culture. *J Orthop Res* 2008;26(10):1306-12.
32. Kawahara E, Oda Y, Katsuda S, Nakanishi I, Aoyama K, Tomita K. Microfilamentous type VI collagen in the hyalinized stroma of the hypertrophied ligamentum flavum. *Virchows Arch A Pathol Anat Histopathol* 1991;419(5):373-80.
33. Grant W, Sullivan R, Sonenshine D, Adam M, Slusser J, Carson K, et al. Electron microscopic investigation of the effects of diabetes mellitus on the Achilles tendon. *J Foot Ankle Surg* 1997;36(4):272-8.